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Canad. J. Bot. 70: 107-113.

REFNO:

2097

KEYWORDS:

Chemistry, Ramonda, Yugoslavia



Effects of dehydration and rehydration on the polar lipid and fatty acid composition of Ramonda species

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Received February 12, 1991

STEVANOVIC, B., PHAM THI, A. T., MONTEIRO DE PAULA, F., and VIEIRA DA SILVA, J. 1992. Effects of dehydration and rehydration on the polar lipid and fatty acid composition of Ramonda species. Can. J. Bot. 70: 107-113.

Endemic Balkan species Ramonda serbica and Ramonda nathaliae (Gesneriaceae) are rare resurrection flowering plants, known to withstand repeated cycles of desiccation—rehydration in their natural habitat. Analysis of their leaf lipids and fatty acids revealed a rather small amount of total lipids (15.8 mg/g dry weight in R. serbica and 19.5 in R. nathaliae) and galactolipids, particularly the monogalactosyl-diacylglycerol, considerably poorer in linolenic acid in comparison to other flowering plants (55% for R. nathaliae and 64% for R. serbica). Severe desiccation leads to a drastic loss in total lipids (76% in R. serbica and 71.5% in R. nathaliae), especially in monogalactosyl-diacylglycerol, but the recovery is extremely rapid and thorough upon the rewetting. A shift towards the more saturated oleic and linoleic acids in galactolipids occurs at a different time and to a different degree in the two species, and it is not evenly repaired. Ramonda serbica, known to be somewhat less resistant, shows a wider range of lipid changes.

Key words: lipid, fatty acids, desiccation-tolerant plants, Ramonda serbica, Ramonda nathaliae.

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Deux espèces endémiques du Balkan, Ramonda serbica et Ramonda nathaliae (Gesnériacées) appartiennent au groupe très restreint des plantes à fleurs reviviscentes. Elles sont capables de supporter, dans les conditions naturelles, des cycles répétés de dessication suivie de réhydratation. L'analyse de leurs lipides foliaires révèle une teneur relativement faible en lipides (15,8 mg par g de matière sèche chez R. serbica et 19,5 chez R. nathaliae) et un pourcentage d'acide linolénique étonnamment bas des galactolipides, en particulier du monogalactosyl-diacylglycérol, comparé aux autres Angiospermes (55% chez R. nathaliae et 64% chez R. serbica). Une déshydratation sévère induit une perte importante des lipides totaux (perte de 71,5% chez R. nathaliae et de 76% chez R. serbica), surtout du monogalactosyl-diacylglycérol. Cependant à la réhydratation, la reprise est extrêmement rapide et totale. La sécheresse provoque une diminution du degré d'insaturation des acides gras des galactolipides, et à la réhydratation, la reprise est seulement partielle. Ramonda serbica, connue pour être moins résistante que R. nathaliae, présente des variations de plus grande amplitude au cours de la déshydratation.

Mots clés: lipide, acides gras, plantes reviviscentes, Ramonda serbica, Ramonda nathaliae.

Introduction

The discovery of Ramonda serbica Panc. (Pancic 1874) (Fig. 1) and Ramonda nathaliae Panc. et Petrov. (Petrovic 1885) (Fig. 2), the endemic relics of the Balkan Peninsula, was met with great interest among the botanists of the time, since those Ramonda species are both typical relics and at the same time, rare European representatives of the tropic-subtropic family, the Gesneriaceae. These resurrection flowering plants belong to the tertiary mountainous flora, and the glacial age may have been the period when the climatic changes forced them towards the lower regions of the Balkans, where they are found nowadays. Their survival in the present day habitat is largely dependent on drought resistance that they achieve through complete desiccation, and Ramonda species are known to enter this state several times during their lifetime. In the process, the plants undergo complete desiccation

ABBREVIATIONS: DGDG, digalactosyl-diacylglycerol; DPG, diphosphatidylglycerol; FAC, fatty acid composition; GL, galactolipids; MGDG, monogalactosyl-diacylglycerol; NL, neutral lipids; PC, phosphatidylcholine; PE, phosphatidylchanolamine; PG, phosphatidylglycerol; PI, phosphatidylinositol; PL, phospholipids.

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(Fig. 3), suspending their metabolism but maintaining the ability to resume full biological activities upon rehydration.

Water deficit could easily be expected to affect cell membranes profoundly through structural changes and possible breakdown. An efficient repair and full reconstitution of membrane integrity during rehydration would be a prerequisite to cell survival.

Polar lipid changes could be anticipated in the process, and the subject has been receiving considerable attention. Fatty acid composition of desiccation tolerant mosses was found to change during water stress as well as the activity of several enzymes involved in lipid metabolism; two different enzymatic pathways were proposed to explain tissue defence against the uncontrolled loss of unsaturated fatty acids (Dhindsa and Matowe 1981; Stewart and Bewley 1982). In higher plants, water deficit leads to a decline in polyunsaturated fatty acid content of leaves, particularly in the glycolipid fractions (Chetal et al. 1981; Pham Thi et al. 1982; Wilson et al. 1987; Monteiro de Paula et al. 1990). It also results in a profound overall drop in MGDG, the major leaf glycolipid (Ferrari-Iliou et al. 1984), owing to an increased lipolytic activity (El-Hafid et al. 1989) and to the inhibition of precursor uptake and desaturation activity (Pham Thi et al. 1985, 1987).

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Fig. 1. A well-hydrated plant of Ramonda nathaliae in its natural habitat



Fig. 2. A well-hydrated plant of Ramonda serbica in its natural habitat.

To date knowledge is lacking on the lipid content and composition of the two *Ramonda* species, and the present study was undertaken to examine them as they appear in restricted water supply conditions, as well as during desiccation and recovery.

Materials and methods

Plant material and drought treatment

Whole plants (including soil surrounding the roots) of *R. serbica* and *R. nathaliae* were collected from their natural habitat in the south eastern regions of Serbia. Yugoslavia. They were transferred to the laboratory and put into flat pots where some plants were kept well hydrated at all times (control plants) while the others were subjected to drought by withholding water. Relative humidity of the air was

kept at 30%. The drought treatment lasted 7 days. The severely desiccated plants were then rehydrated by spraying distilled water on the leaves and by rewatering the soil.

Lipid and fatty acid analyses were made on the following plants: (i) control plants (C): water potential $(\psi) = -0.5$ to -0.6 MPa. WC = 80%; (ii) moderately stressed plants (S1, 4 days dehydration): $\psi = -1.9$ to -2.0 MPa. WC = 53%; (iii) severely stressed plants (S2, 7 days dehydration): $\psi =$ unmeasurable (less than -6 MPa). WC = 11%; (iv) plants rehydrated for 2 days (R1): $\psi = -0.6$ to -0.7 MPa, WC = 77.5%; and (v) plants rehydrated for 6 days (R2) $\psi = -0.5$ to -0.6 MPa, WC = 82%.

Water potentials (MPa) were measured in a pressure chamber (Scholander et al. 1964). The water content (WC) is the percentage of water on a fresh weight basis and equals the fresh weight minus the dry weight divided by the fresh weight and multiplied by 100

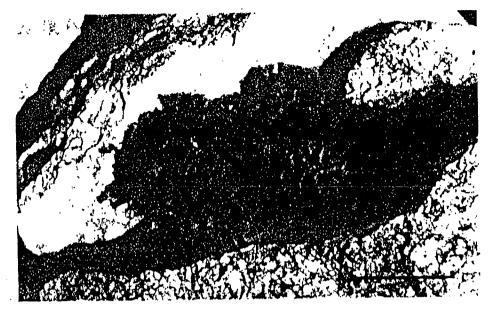


Fig. 3. Desiccated plants of Ramonda in natural habitat.

Limd analysis

The leaves were fixed in boiling water and lipids were extracted in chloroform—methanol (Allen and Good 1971). Lipids were separated by thin layer chromatography (Lepage 1967). Fatty acids from the total lipid extracts and from the lipid classes were quantified by gas—liquid chromatography, using heptadecanoic acid as internal standard rfor details of the methods, see Pham Thi et al. 1985).

All results are means of three replicates \pm SD.

Results

Total leaf lipid content

The polar lipid content of leaves was found to be in the range of 15 to 20 mg/g DW for both Ramonda species (Fig. 4), a rather low value compared with that usually found in other flowering plants (30 to 50 mg/g DW) (see for example Pham Thi et al. 1990). Progressive desiccation resulted in a steady and steep reduction, so that in severely stressed plants it was well below the half of the initial values. This profound drop showed an extremely rapid and thorough recovery upon rewatering, and in 2 days the lipid content was regained, rising even somewhat over the original values. This pattern was similar in both species.

Fatty acid composition of total leaf lipids

The analysis showed the lipids to consist of palmitic (C 16:0), palmitoyl (C 16:1), stearic (C 18:0), oleic (C 18:1). linoleic (C 18:2), and linolenic (C 18:3) acids (Table 1), as found in the majority of higher plants, particularly those who belong to the so-called 18:3 group (Jamieson and Reid 1971). It is noteworthy that the main component is linoleic acid, which accounted for about 50% of the total fatty acid content, whereas linolenic acid represented about 30% in contrast with other flowering plants in which linolenic acid is the most abundant fatty acid in the green leaves (see Benson 1964).

In *R. nathaliae*, desiccation led to little changes in the relative distribution of the main fatty acids, except a slight increase in C 18:3 percentage under moderate stress, paralleled to some extent by a decrease in C 18:2. In *R. serbica*, in severely stressed leaves, the C 18:3 percentage decreased significantly, while that of C 18:2 increased.

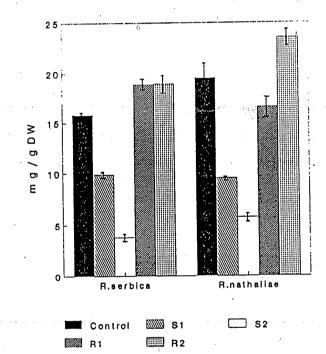


Fig. 4. Effects of dehydration and rehydration on the total lipid content of *Ramonda* leaves (in mg/g DW). Control, well-hydrated plants; S1, moderately stressed plants; S2, severely stressed plants; R1, plants rehydrated for 2 days; R2, plants rehydrated for 6 days. For more details, see Materials and methods.

Following rehydration, the relative proportion of C 18:3 to C 18:2 did not regain its normal values, even after 6 days.

Lipid composition

In Figs. 5 and 6 the main lipid classes of *Ramonda* leaves are represented in milligrams per gram leaf dry weight. In normal conditions, the galactolipid MGDG appeared as the major lipid class. In both species it was greatly reduced in severe desiccation but nearly fully recovered early during rehydration. While having roughly the same general pattern of lipid

TABLE 1. Effect of dehydration and rehydration on the fatty acid composition of total lipids in Ramonda leaves

	Fatty acid (weight as % of total lipids)							
Treatment*	. 16:0	16 : 1	18:0	18:1	18:2	18:3		
		Pam	onda natha	lina	ţ)			
		Name	maa nama	iitie				
С	11.0 ± 0.4	2.1	1.9	2.4	50.2 ± 2.4	32.3 ± 2.5		
Šì	8.9 ± 0.9	1.9	2.1	3.4	47.4 + 2.5	36.2 ± 1.8		
S2	11.2 ± 0.5	1.2	0.9	1.7	51.2 ± 1.9	33.6 ± 2.0		
RI	11.3 ± 1.3	3.0	2.7	4.3	53.1 ± 1.7	25.7 ± 0.8		
R2	12.4±0.8	2.1	1.7	4.8	49.3±1.5	29.7±1.1		
		Ran	nonda serb	ica				
С	11.5 ± 0.7	1.9	1.9	2.8	48.4 ± 2.3	33.4 ± 3.3		
SI	11.4 ± 1.5	3.9	2.1	4.2	46.1 ± 3.2	32.3 ± 2.8		
S2	11.9±0.9	1.2	1.6	4.6	54.0±2.9	26.7 ± 3.3		
	_	2.1	1.0	3.1	53.6±1.0	29.2 ± 0.9		
RI	10.8±2.1		2.1	5.6	52.3 ± 2.6	25.2 ± 1.6		
'R2	12.3±1.7	2.6	۷,۱	J.0	32.3 ±2.0	25.211.0		

^{*}C, control, well-hydrated plants; \$1, plants submitted to moderate water stress; \$2, plants submitted to severe water stress; R1, plants rehydrated for 2 days; S2, plants rehydrated for 6 days.

class distribution, the two Ramonda species showed certain differences during water stress.

In R. serbica (Fig. 5), the drop in MGDG content was extremely severe before the very rapid and nearly complete recovery upon rewetting. The same trend in changes, but to a much lesser extent, was observed in DGDG and in PC. In contrast, PE, other phospholipids (including PI, PG, and DPG), and neutral lipids (NL) were much less touched by desiccation. In fact, their relative amount rose remarkably during desiccation to triple and double that of the initial levels (23.7% of total lipids in S2 leaves instead of 7% in C for PE, 20.4% instead of 8.8% for the other phospholipids, and 21.1% instead of 12.7% for NL).

In R. nathaliae (Fig. 6), the drop in MGDG content under drought stress was also severe, but much less than in R. serbica. On the other hand, the decrease in PE and NL was more important. Consequently, in desiccated leaves, the relative distribution of lipid classes is closer to that in control leaves, suggesting a greater stability of the membranes in R. nathaliae than in R. serbica. On rehydration, the situation rapidly became normal.

Fatty acid composition of lipid classes

The fatty acid composition of the four main lipid classes (MGDG, DGDG, PC, and PE) is presented in Table 2 (R. serbica) and Table 3 (R. nathaliae).

Linolenic acid (18:3) was found to be the main fatty acid component in MGDG, but in a lower percentage, i.e., 55-65% compared with 80-90% in the typical flowering plants (Douce and Joyard 1980). Water stress decreased its content; it reached the lowest point during severe desiccation in R. serbica leaves (Table 2) and in early rehydration in R. nathaliae leaves (Table 3). Again, the drop was much more pronounced in R. serbica (from 64.4 to 29.7%), where the percentage also remained low into the rehydration period. The loss of linolenic acid during water stress was closely accompanied by the rise in linoleic acid (18:2) levels.

Linolenic and linoleic acids were nearly equally represented in DGDG composition. The similar shift towards the less unsaturated fatty acid was observed to a various degree in DGDG of both species during water stress.

Concerning the phospholipids PC and PE, the relative distribution of palmitic (C 16:0), linoleic (C 18:2), and linolenic (C 18:3) acids depends on the intensity of the stress as well as on the plant species. Thus, in R. nathaliae, the unsaturation of PC and PE increased under stress, owing to an increase in the ratio of C18:3 to C18:2, whereas in R. serbica it decreased. Upon rehydration, the fatty acid composition of PC and PE did not return to the control values.

Discussion

Resurrection flowering plants are rare in the existing world vegetation, and nearly all their representatives occur in the southern hemisphere (Gaff 1977, 1981). Endemic species R. serbica and R. nathaliae are in this respect all the more unusual since they occur in the northern hemisphere. Very similar in appearance, their taxonomic validity was challenged soon after their discovery (Velenovsky 1898; Vandas 1909; Doflein 1921). Soon enough, their distribution and morphological and ecological differences (Kosanin 1939), along with chromosome specificities (Glisic 1924), established them as independent sibling species. More recent investigations on the respective areas of distribution and their climatic and geographic properties (Stevanovic and Stevanovic 1985; Stevanovic et al. 1986a, 1987), and particularly on the ecophysiological characters and water balance of both species (Stevanovic 1986). clearly demonstrated that the two species show noticeably different preferences for habitat conditions. Although both are successful resurrection plants, the somewhat more xeromorphic R. nathaliae also appears to be more resistant to harsh climate. This slightly different tolerance is maintained even in rare sympatric habitats (Stevanovic et al. 1986b) where it is translated into recognizably specific microenvironments.

Grossly changed appearance under desiccation with evident overall plant friability is highly suggestive of potentially serious structural damage. Nevertheless, rewatering brings about a rapid and thorough recovery of full biological potential in both Ramonda species. Little general knowledge is available on the composition of structural lipids in resurrection flowering plants. In this regard, lipid analysis of Ramonda species

brought to light several distinctive features.

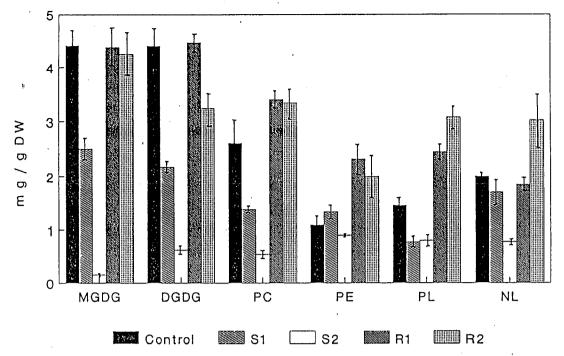


Fig. 5. Effects of dehydration and rehydration on the lipid content of *Ramonda serbica* leaves (in mg/g DW). MGDG, monogalactosyldiacylglycerol; DGDG, digalactosyl-diacylglycerol; PC, phosphatidyl-choline; PE, phosphatidyl-ethanolamine; PL, other phospholipids; NL, neutral lipids. Other abbreviations as in Fig. 4.

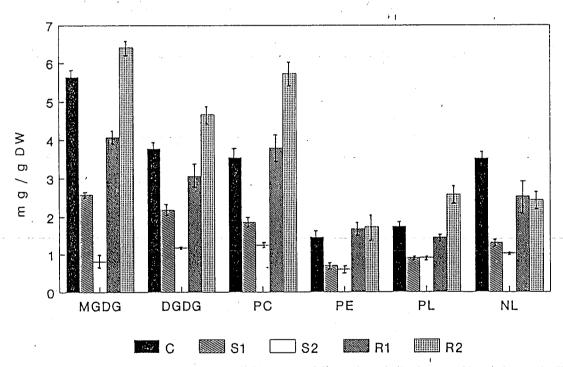


Fig. 6. Effects of dehydration and rehydration on the lipid content of Ramonda nathaliae leaves. Abbreviations as in Fig. 5.

The leaves of *Ramonda* species are rather low in total lipids in comparison to other flowering plants (see for example Pham Thi et al. 1990) and this suggests a low quantity of membrane material. Another unexpected feature is a peculiarly low content of linolenic acid in galactolipids, i.e., 54.7% in MGDG from *R. serbica* and 64.4% in MGDG from *R. nathaliae* instead of 90-95% in other flowering plants (see Benson 1964).

Glycolipids containing high percentages of polyunsaturated fatty acids are characteristic of chloroplast membranes (Douce and Joyard 1980).

It is generally accepted that membrane fluidity, i.e., the degree of order and motion in the hydrocarbon core of the lipid bilayer (Stubbs and Smith 1984), depends on the degree of unsaturation of the faity acids (Stubbs and Smith 1984;

TABLE 2. Effects of dehydration and rehydration on the fatty acid composition of the main lipid classes from Ramonda serbica leaves

Fatty acid (weight as % of total lipids) Lipid and treatment* 16:0 18:0 18:2 18:1 18: 3 MGDG C 1.2 2.6 1.0 30.8 ± 0.3 64.4 ± 1.9 SI 1.4 2.4 2.0 37.4 ± 0.5 56.8 ± 2.1 52 8.5 0.4 16.0 45.4 ± 1.5 29.7±0.7 R1 1.3 1.6 2.1 48.9 ± 1.0 46.1 ± 0.4 R2 2.6 2.1 . 2.3 55.7 ± 1.9 36.3 ± 1.7 DGDG C 7.1 0.1 1.5 38.8 ± 3.2 52.5 ± 3.0 SI 5.9 0.6 1.1 48.1 ± 1.1 44.2 ± 1.4 S2 6.6 0.2 1.2 54.0 ± 2.3 38.2 ± 2.5 R! 3.4 1.6 0.9 53.1 ± 0.6 41.0 ± 1.7 R2 6.1 1.6 0.9 55.2 ± 0.8 36.2 ± 1.3 PC C 18.9 ± 1.3 3.6 2.7 55.8 ± 0.3 19.0 ± 1.2 SI 4.2 16.6±1.6 4.0 61.1 ± 1.9 14.1 ± 0.9 S2 16.6 ± 1.4 10.6 3.4 56.9±1.8 12.5 ± 1.8 RΙ 15.9 ± 1.3 3.7 6.3 59.1 ± 1.3 15.0 ± 0.4 R2 11.9 ± 1.7 1.3 6.7 64.8 ± 1.9 15.3 ± 0.3 PΕ C 24.3 ± 2.1 3.9 4.1 58.8 ± 3.2 8.9 ± 2.8 SI 25.6 ± 2.5 2.0 7.8 54.9 ± 0.8 9.7 ± 0.2 S2 22.8 ± 1.0 7.6 ± 1.0 6.1 11.0 52.5 ± 0.4 R1 25.1 ± 3.8 4.7 7.4 53.0 ± 3.9 9.8 ± 1.2 R2 20.8 ± 0.7 0.8 9.9 57.2±3.7 11.3 ± 0.5

Bishop et al. 1979). Concerning chloroplast membranes, the efficiency of photosynthesis seems to be largely dependent on their fluidity (Bishop et al. 1982; Ford and Barber 1983). Therefore, the net effect of the observed low linolenic acid content would be a stabilized bilayer and a decreased overall fluidity of the chloroplast membrane and hence its metabolic activity. In resistant membranes, we hypothesize that the price to pay for a greater stability is a reduced efficiency (Pham Thi et al. 1990).

Under water stress, polar lipid content of Ramonda leaves decreased, attesting a great loss of membrane; the galactolipid MGDG is the most susceptible to degradative processes induced by drought. A similar phenomenon has been observed in several other plants (Chetal et al. 1981; Ferrari-Iliou et al. 1984; Monteiro de Paula et al. 1990). Finally, the lipid composition of desiccated Ramonda leaves is profoundly modified: the ratio of PL to GL increased and the relative proportion of MGDG to DGDG drastically decreased. An increase in the phospholipids relative to galactolipids in leaves indicate a preferential degradation of the chloroplast membranes, and in some stress conditions, at dawn for example, the ratio of MGDG to DGDG decreased as well as the degree of unsaturation of galactolipids, as shown by Öquist (1982).

In vitro, pure lipids spontaneously form bilayers. However, depending on hydration and temperature, they can adopt different configurations (see Seddon 1990). It has been shown that MGDG frequently forms the HexII phase whereas DGDG adopts only lamellar phases. PE has a strong tendency to form the HexII phase, while PC does not form any nonlamellar phases (see Seddon 1990). Even if natural membranes are

TABLE 3. Effects of dehydration and rehydration on the fatty acid composition of the main lipid classes from *Ramonda nathaliae* leaves

Lipid and	Fatty acid (weight as % of total lipids)						
treatment*	16:0	18:0	18:1	18:2	18: 3		
MGDG	ŀ						
C	1.2	1.3	1.1	41.7 ± 0.7	54.7±0.6		
SI	1.0	_	0.1	40.1 ± 0.7	57.9 ± 1.6		
S2	2.8		4.2	42.0 ± 0.8	51.0 ± 1.7		
RI	2.2	0.4	3.1	52.8 ± 1.9	41.5 ± 0.6		
R2	1.7	1.3 '	4.9	40.4 ± 1.7	51.7 ± 1.3		
DGDG					. –		
C ''	4.3	3.2	2.0	46.5 ± 0.8	44.0 ± 0.6		
SI	5.8	0.6	2.9	47.7 ± 0.6	43.0 ± 0.5		
S2	6.8	5.3	2.3	46.2 ± 1.3	39.4 ± 0.3		
RI	7.4	4.7	3.7	53.4 ± 1.6	30.8 ± 2.5		
R2	5.1	0.4	1.9	50.5 ± 0.9	42.1 ± 1.3		
PC					_		
С	16.6 ± 1.2	2.5	3.1	63.9 ± 0.8	13.9 ± 0.9		
S1	14.3 ± 0.4	3.8	3.4	59.7 ± 0.3	18.8 ± 0.7		
S2	14.1 ± 0.6	8.9	5.7	52.9 ± 2.5	18.4 ± 0.5		
RI ·	16.9 ± 0.2	3.8	6.0	57.2 ± 2.6	16.1 ± 0.3		
R2	16.4 ± 0.8	5.7	6.7	54.5 ± 0.7	16.7 ± 0.5		
PE					(
C ·	20.5 ± 1.4	2.0	2.5	66.7+0.7	8.3 ± 0.3		
SI	15.2±0.9	2.5	7.8	62.2 ± 1.8	12.3 ± 1.5		
S2	20.4 ± 1.7	8.6	8.2	52.9±0.9	9.9 ± 1.6		
RI	21.0 ± 3.0	4.4	6.5	57.6 ± 1.9	10.5 ± 1.1		
R2	12.0±0.8	6.4	4.9	66.4 ± 2.2	10.3 ± 2.3		

^{*}See Table 1.

complex mixtures of several lipid classes, it is not doubtful that variations in their relative proportions induced by drought could influence the physical state of the membrane (Gordon-Kamm and Steponkus 1984; Pearce 1985; Norberg et al. 1990) and hence the activity of membrane proteins (Spector and Yorek 1985).

Though the reduction of *Ramonda* leaf lipids was drastic under dehydration, synthesis upon rewatering is strikingly fast. Similar rapid and extensive changes in phospholipids are observed in drought-tolerant mosses (Stewart and Bewley 1982). Investigations on mosses as well as on other resurrection angiosperms (e.g., *Xerophytica* sp.) report a great stability of the mRNA under desiccation and a rapid synthesis of polyribosomes and proteins on rehydration (Tymms and Gaff 1984; Tucker and Bewley 1976; Dhindsa and Bewley 1978).

The comparison of lipid modifications in the two species is particularly interesting. Ramonda serbica, known to prefer drier habitats than R. nathaliae (Stevanovic et al. 1987), also shows a greater stability of the membrane lipids and a better capacity to resume on rewatering, particularly regarding fatty acid composition. Clearly, the lipid analysis offers a new insight into the survival strategies of Ramonda sp. The observed differences between the two resistant species support previous knowledge of ecological features (Stevanovic 1986) and signal the possibility that specific adaptations might be used to ensure their respective water stress tolerance.

Allen, C. F., and Good, P. 1971. Acyl lipids in photosynthetic systems. Methods Enzymol. 23: 533-547.

Benson, A. A. 1964. Plant membrane lipids. Annu. Rev. Plant Physiol. 15: 1-16.

^{*}See Table 1.

Bishop, D. G., Kenrick, J. R., Bayston, J. H., MacPherson, A. S., Johns, S. R., and Willing, R. I. 1979. The influence of fatty acid unsaturation on fluidity and molecular packing of chloroplast membrane lipids. *In* Low temperature stress in crop plants. The role of the membrane. *Edited by J. M. Lyons*, D. Graham, and J. K. Raison. Academic Press, New York. pp. 375-390.

Bishop, D. G., Kenrick, J. R., Coddington, J. M., Johns, S. R., and Willing, R. I. 1982. The role of membrane fluidity in the maintenance of chloroplast function. *In Biochemistry and metabolism of plant lipids. Edited by J. F. G. M. Wintermans and P. J. C. Vicinia and P. J. C. William and P. J. C.*

Kuiper. Elsevier, Amsterdam. pp. 339-344.

Chetal, S., Wagle, D. S., and Nainawatee, H. S. 1981. Glycolipid changes in wheat and barley chloroplast under water stress. Plant Sci. Lett. 20: 225-230.

- Dhindsa, R. S., and Bewley, J. D. 1978. Messenger RNA is conserved during drying of the drought-tolerant moss *Tortula ruralis*. Proc. Natl. Acad. Sci. U.S.A. 75: 842-846.
- Dhindsa, R. S., and Matowe, W. 1981. Drought tolerance in two mosses is correlated with enzymatic defence against lipid peroxidation. J. Exp. Bot. 32: 79-91.
- Doflein, F. 1921. Mazedonien, Erlebnisse und Beobachtungen eines Naturfoschers in Gefolge des Deutschen Heeres. G. Fischer, Jena.
- Douce, R., and Joyard, J. 1980. Chloroplast envelope lipids: detection and biosynthesis. Methods Enzymol. 69: 290-301.
- El-Hafid, L., Pham Thi, A. T., Zuily-Fodil, Y., and Vieira da Silva, J. 1989. Enzymatic breakdown of polar lipids in cotton leaves under water stress. I. Degradation of monogalactosyl-diacylglycerol. Plant Physiol. Biochem. 27: 495-502.
- Ferrari-Iliou, R., Pham Thi, A. T., and Vieira da Silva, J. 1984. Effect of water stress on the lipid and fatty acid composition of cotton (Gossypium hirsutum L.) chloroplasts. Physiol. Plant. 62: 219-224.
- Ford, R. C., and Barber, J. 1983. Incorporation of sterol into chloroplast thylakoid membranes and its effect on fluidity and function. Planta, 158: 35-41.
- Gaff, D. F. 1977. Desiccation tolerant vascular plants of Southern Africa. Oecologia, 31: 95-105.
- Gaff, D. F. 1981. The biology of resurrection plants. *In* The biology of Australian plants. *Edited by J. S. Pate and A. J. McComb. University of Western Australia Press, Perth. pp. 114-146.*
- Glisic, L. J. 1924. Development of the female x generation and embryo in Ramonda. Ph.D. thesis, University of Belgrade, Belgrade, Yugoslavia.
- Gordon-Kamm, W. J., and Steponkus, P. L. 1984. Lamellar-to-hexagonal II phase transitions in the plasma membrane of isolated protoplasts after freeze-induced dehydration. Proc. Natl. Acad. Sci. U.S.A. 81: 6373-6377.
- Jamieson, G. R., and Reid, E. H. 1971. The occurrence of hexadeca-7,10,13-trienoic acid in the leaf lipids of angiosperms. Phytochemistry, 10: 1837-1843.
- Kosanin, N. 1939. Gradja za biologiju Ramondia Nathaliae, R. Serbica i Ceterach officinarum. Spom. Srp. Kral Akad. 89: 1-68.
- Lepage, M. 1967. Identification and composition of turnip root lipids. Lipids, 2: 244-250.
- Monteiro de Paula, F., Pham Thi, A. T., Vieira da Silva, J., Justin, A. M., Demandre, C., and Mazliak, P. 1990. Effects of water stress on the molecular species composition of polar lipids from Vigna unguiculata leaves. Plant Sci. 66: 185-193.

Norberg, P., Larsson, K., and Liljenberg, C. 1990. A study of membrane lipids from dehydration-acclimated *Brassica napus* root cells: formation of a cubic phase under physiological conditions. Biochem. Cell Biol. 68: 102-105.

- Öquist, G. 1982. Seasonally-induced changes in acyl lipids and fatty acids of chloroplast thylakoids of *Pinus silvestris*. A correlation between the level of unsaturation of monogalactosyldiglyceride and the rate of electron transport. Plant Physiol. 69: 869-875.
- Pancic, J. 1874. Flora knezevine Srbije. Belgrade, Yugoslavia. Pearce, R. S. 1985. The membranes of slowly drought-street.
- Pearce, R. S. 1985. The membranes of slowly drought-stressed wheat seedlings: a freeze-fracture study. Planta, 166: 1-14.

- Petrovic, S. 1885. Ramondi je u Srbiji. Glas. Srp. Ucen. Drus. 62: 101-123.
- Pham Thi, A. T., Flood, C., and Vieira da Silva, J. 1982. Effects of water stress on lipid and fatty acid composition of cotton leaves. *In* Biochemistry and metabolism of plant lipids. *Edited by J. F. G. M.* Wintermans and P. J. C. Kuiper. Elsevier, Amsterdam. pp. 451–454.
- Pham Thi, A. T., Borrel-Flood, C., Vieira da Silva, J., Justin, A. M., and Mazliak, P. 1985. Effects of water stress on lipid metabolism in cotton leaves. Phytochemistry, 24: 723-727.
- Pham Thi, A. T., Borrel-Flood, C., Vieira da Silva, J., Justin, A. M., and Mazliak, P. 1987. Effects of drought on [1-14C]-oleic and [1-14C]-linoleic acid desaturation in cotton leaves. Physiol. Plant. 69: 157-159.
- Pham Thi, A. T., Vieira da Silva, J., and Mazliak, P. 1990. The role of membrane lipids in drought resistance of plants. Bull. Soc. Bot. Fr. 137: 99-104.
- Scholander, F., Hammel, H. T., Hemmingsen, E. A., and Bradstreet, E. D. 1964. Hydrostatic pressure and osmotic potential in leaves of mangrove and some other plants. Proc. Natl. Acad. Sci. U.S.A. 52: 119-125.
- Seddon, J. M. 1990. Structure of the inverted hexagonal (HII) phase, and nonlamellar phase transitions of lipids. Biochim. Biophys. Acta, 1031: 1-69.
- Spector, A. A., and Yorek, M. A. 1985. Membrane lipid composition and cellular function. J. Lipid Res. 26: 1015-1035.
- Stevanovic, B. 1986. Ecophysiological characteristics of the species Ramonda serbica Panc. and R. nathaliae Panc. et Petrov. Ekologija (Belgrade), 21: 119-134.
- Stevanovic, V., and Stevanovic, B. 1985. Asplenio cuneifolii and Ramondaetum nathaliae new chasmophytic community on serpentine rocks in Macedonia. Glas. Prir. Muz. Bcogr. Ser. A. 40: 75-87.
- Stevanovic, V., Niketic, M., and Stevanovic, B. 1986a. On distribution of endemic and relic species Ramonda serbica Panc. (Gesneriaceae) in SR Macedonia (Yugoslava). Glas Prir. Muz. Beogr. Ser. A, 41: 17-25.
- Stevanovic, V., Niketic, M., and Stevanovic, B. 1986b. Sympatric area of the sibling and endemo-relic species Ramonda serbica Panc. and R. nathaliae Panc. et Petrov. (Gesneriaceae) in southeast Serbia (Yugoslavia). Glas. Inst. Bot. Univ. Beogr. 20: 45-54.
- Stevanovic, V., Niketic, M., and Stevanovic, B. 1987. Phytocoenological characteristics of sympatric habitats of endemo-relic species *Ramonda serbica* Panc. and *R. nathaliae* Panc. et Petrov. Glas. Inst. Bot. Univ. Beogr. 21: 17-26.
- Stewart, R. C., and Bewley, J. D. 1982. Stability and synthesis of phospholipids during desiccation and rehydration of a desiccationtolerant and a desiccation-intolerant moss. Plant Physiol. 69: 724-727.
- Stubbs, C. D., and Smith, A. D. 1984. The modification of mammalian polyunsaturated fatty acid composition in relation to membrane fluidity and function. Biochim. Biophys. Acta, 779: 89-137.
- Tucker, E. B., and Bewley, J. D. 1976. Plant desiccation and protein synthesis. III. Stability of cytoplasmic RNA during dehydration and its synthesis on rehydration of the moss *Tortula ruralis*. Plant Physiol. 57: 564-567.
- Tymms, M. J., and Gaff, D. F. 1984. Recovery of protein synthesis and RNA synthesis after rewetting of desiccated leaves of the Angiosperm *Xerophytica villosa*. Biochem. Physiol. Pflanz. 179: 211-217.
- Vandas, C. 1909. Reliquiae Formanekinae. Brunae.
- Velenovsky, J. 1898. Supplementum florae bulgaricae. Prague.
- Wilson, R. F., Burke, J. J., and Quisenberry, J. E. 1987. Plant morphological and biochemical responses to field water deficit: responses of leaf glycerolipid composition in cotton. Plant Physiol. 84: 251-254.